Algae ATX: Molecular and Biochemical Screening for Toxic Algae, City of Austin, Texas

Project proposal prepared by Dr. Schonna R. Manning Assistant Professor | Department of Biological Sciences Omics Coordinator | Institute of Environment Florida International University – Biscayne Bay

1.0 Objectives

- 1.1 The purpose of this proposal is to expand the use of analytical techniques available to screen for toxins and identify the algal species that produce toxins in our drinking and recreational water supplies. After the 2019 harmful algal bloom event, an increased need is recognized to track the development of a bloom and identify and quantify algal toxins present that can negatively impact recreational uses of Austin's reservoirs. Early detection through monitoring will enable more accurate prediction of harmful bloom development and targeted management strategies toward mitigation, and hopefully, prevention. This work supports City of Austin programs to monitor waterways for nuisance algal species that may result in impairments of the beneficial uses desired by the community.
- 1.2 Objectives of the proposed work are to assist the City of Austin in the following areas:
 - 1.2.1 Expand our understanding and description of the algae species present in our streams and reservoirs by combining traditional taxonomic identifications with DNA extraction and sequencing;
 - 1.2.2 Develop a molecular library for identification of dominant or problematic species and weather those species have the potential to produce toxins of concern;
 - 1.2.3 Identify and quantify toxin(s) present in algal cells, mats, and/or water samples. Toxin(s) present will be linked to specific cyanobacterial species through molecular fingerprinting results

2.0 Background

- 2.1 Cyanobacteria blooms have been recognized as a significant emerging threat to municipal and recreational water supplies. Under certain environmental conditions, the blooms have been found to produce toxins that have been linked to skin rashes, cancer, and even death of those that come into contact with or consumes impacted waters. It is therefore essential to monitor water supplies for the presence of or potential for toxigenic cyanobacteria to impair aquatic systems. In the Lake Austin reservoir during a recent drought of record, cyanobacteria bloom magnitude and duration significantly increased, leading to concerns about the potential for the reservoir that is a drinking water source as well as popular recreational reservoir to become impacted by cyanotoxins (Bellinger et al. 2018).
- 2.2 Because of the increased cyanobacteria biomass, a pilot study identified phytoplankton

species during blooms and screened the water for toxins (Bellinger 2018). That study did not find the presence of toxins in the water. However, it was speculated that some species may have the capability to produce toxins. This uncertainty in the potential for the species present to produce toxins led staff to seek out additional means of determining the risk of the species present to the reservoirs. In addition, the study was limited to only Lake Austin and Lady Bird Lake.

- 2.3 Based on the desire to further understand the potential for species to produce toxins as well as expand the screening of Austin's important aquatic systems, The City is looking to contract with the University of Texas which has developed the methods and techniques to isolate species of interest, analyze their DNA for the potential to produce toxins, and apply marker screening to rapidly determine if the species of concern is present in the aquatic system of interest.
- 2.4 In 2019, a distinctive cyanobacteria bloom event, consisting of cohesive mats that grow on the sediment surface and float at the surface after becoming buoyant, occurred in the upper half of the Lady Bird Lake reservoir, notably around Red Bud Isle and Auditorium shores. The harmful bloom event resulted in the death of multiple dogs after likely ingestion of algal biomass from contact with the mats. It was determined that a neurotoxin was present in the mats, with impacts consistent with the symptoms displayed by the dogs prior to their passing.
- 2.5 In light of the harmful bloom event, the scope of the previous pilot project is being expanded to build on the initial findings but to now include monitoring of cohesive cyanobacterial mats and to have toxin identification and quantitation provided in concert with the molecular screening.

3.0 Scope Of Work, 2023

The Manning Laboratory at FIU-BBC will continue generate to biochemical profiling and quantitative analysis of targeted algal toxins. These biochemical characteristics will be added to the micrographic and barcoding library for the rapid assessment of toxic algae in Austin waterways. The examination of benthic strains of microalgae will be added to these analyses, which was previously monitoring phytoplankton, exclusively.

The Manning Lab will generate micrographic and genetic barcoding libraries to develop a reference guide for the identification of microalgae strains (primarily cyanobacteria) that occur in Austin waterways. In addition to DNA barcoding, i.e., 23S, cyanoITS, ITS2, will be used for the identification of microalgal isolates from surrounding waterways.

The 2023 phase of Algae ATX will involve the extraction of environmental DNA (eDNA) to examine the complexity of algal metaphyton mats and community structure. Total eDNA will be extracted from metaphyton mat materials and the DNA will be sequenced using Illumina NovaSeq PE150 whole-genome shotgun sequencing and compared with existing databases for the determination of species diversity, presence and abundance.

3.1 <u>Toxin identification and quantification.</u> Based on the harmful algal bloom of 2019, the need has arisen for rapid, accurate identification and quantitation of toxins being produced by algae. Presence or absence of specific toxins will be linked to the genetic potential of the strain, described in the above work, to produce toxins of concern.

3.1.1 Deliverables: Reports on the suite of toxins present, expressed as concentration (i.e., mass per unit volume of water) or content (i.e., mass per unit dry mass of algae).

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- 3.2 <u>Development of robust molecular tools for the detection and identification of toxin-forming species</u>: In addition to DNA barcoding investigations (e.g., 16S, 23S, ITS2, and toxin-related genes), examination of mat materials have revealed the complexity of the metaphyton mats containing toxigenic microalgae. Initial phases of this project have isolated individual algae for DNA barcoding analysis, although not all isolates could be maintained in culture and/or were overgrown by competing strains. Thus, we propose to develop tools for the holistic analysis of metaphyton biomass, including the microbiome (phycosphere), using environmental DNA (eDNA) and genome sequencing to examine the algal community structure, including species presence/absence and relative abundance.
 - 3.2.1 Deliverables: Reports on DNA barcoding sequences for successful isolates.

3.2.2 Deliverables: Reports on the algal community composition at the different sampling sites, at various timepoints of the monitoring season.

4.0 Tasks

Task 1. Toxin identification and quantitation (2020-2025)

Objectives: Gas chromatography (GC) and/or liquid chromatography (LC) coupled with mass spectrometry (MS) will be utilized to identify and determine concentrations of toxins extracted from water or cyanobacterial samples collected from Austin's reservoirs. Ongoing Task.

- ➔ From 2020 2022, toxin analysis reports were provided weekly to City scientists for algal and water samples collected during the summer-fall months. This dataset is maintained as a real-time Excel spreadsheet for each sample site. Some of the toxin results have been published by peer-reviewed journals, e.g., Manning et al. 2020, and forthcoming manuscripts are planned for the other years, including pre- and post-Phoslock treatments to determine the effectiveness of these treatments on algal abundance and toxin production.
- → This task will continue at FIU for the project years 2023 2025.

Task 2. Phytoplankton cyanobacterial species identification, DNA barcoding analysis (2020-2025)

Objectives: Continue study of applying molecular techniques toward rapid, certain determination of the presence of toxigenic cyanobacteria species in the water column of Austin's reservoirs. Ongoing Task.

- ➔ From 2020 2022, individual taxa from metaphyton materials were isolated for genetic barcoding and DNA sequencing. These data are available to City scientists through a shared cloud folder. Initial results were published in Manning, Perri, and Bellinger 2020 (Data in Brief), Manning et al. 2020 (Mendeley Data), and more recent data obtained will be published (manuscript in preparation, Perri, Bellinger and Manning et al. 2022, Toxins).
- → This task will continue at FIU for the project years 2023 2025.

Task 3. Analysis of metaphyton eDNA to determine algal community composition (2020-2025)

Objectives: Apply molecular, sequencing, and computational techniques to analyze metaphyton eDNA from complex metaphyton material to determine the relative abundance of microbial taxa. Ongoing task.

➔ Following the 2019 sampling season, sequencing environmental DNA (eDNA) has been the preferred modality over single-cell isolations and sequencing, and metaphyton samples obtained from 2020 and 2021 have been processed using eDNA techniques to capture the community structure using metagenetics. These data have been reported to City scientists and the results of these analyses have been presented at national scientific meetings, i.e., PSA 2021 and JASM 2022. Moreover, we anticipate a portion of these data will be published in 2023 (manuscript in preparation, Manning, Rouzbahani, and Bellinger et al.

2023 UPDATED SOW

- 2023, Toxins special issue on the impacts of algal toxins in freshwater systems).
- → This task will continue at FIU for the project years 2023 2025.

Funding:	FY 2023	\$99,910
-	FY 2024	\$99,910
	FY 2025	\$49,517.77 (Jan 1 – March, 30, 2025)

5.0 Cost Section

Cost Rates:

A total of \$99,910 per year is budgeted to cover personnel, materials and supplies, core facilities, and indirect costs for the **Algae ATX** project.

The PI, Dr. Schonna R. Manning, will contribute 1 month of time per year (\$9251.27) to assist with the project goals and deliverables. A postdoctoral associate will be appointed 50% time (\$27,500) to help with the isolation and screening efforts (e.g., genetic and biochemical analyses) as well as data management. Another \$14,208.04 is budgeted for fringe benefits (calculated at 38.66%).

A total of \$6,000 is budgeted to cover materials and supplies, including, but not limited to: nitrile gloves, micropipette tips, glass Pasteur pipets, general reagents, agarose, agar, petri dishes, Thermo GeneClean (up to 5 large kits), Thermo GeneJet (up to 5 large kits), 500- and 1.5-mL polypropylene microfuge tubes, thin-wall PCR tubes, 96-well PCR plates, 6- and 12-well plates for cultures, and PCR primers (i.e., forward and reverse primers for 23S, 18S, ITS, anatoxin, microcystin, and strain-specific detection). Note that all materials and supplies will be purchased through FIU vendors using state educational rates. Another \$780 is budgeted for publication costs to disseminate the research findings.

A total of \$10,000 is budgeted to cover sequencing facility charges These fees include Sanger sequencing for DNA barcoding and related PCR-based sequencing analyses (up to 625 total sequences, including forward and reverse reactions at \$8 per sequence; \$10,000 total). This will also support Next Generation Sequencing (NGS) of whole genomes for targeted algae (up to 10 strains, approx. \$500 per genome, depending on the size of the genome; \$10,000 total).

Per the FIU faculty-staff union by-laws, there is an instituted 3% salary increment per year, thus materials and supplies and publication costs were decreased in subsequent project to accommodate this increase and to stay within the pre-approved budget. The indirect costs are calculated at the current rate of 47.5%.

6.0 Term of Contract

The Contract shall commence upon execution, unless otherwise specified, and shall remain in effect for an initial term of twelve (12) months. The Contract may be extended beyond the initial term for up to four (4) additional twelve (12) month periods at the City's sole option. If the City exercises any extension option, all terms, conditions, and provisions of the Contract shall remain in effect for that extension period, subject only to any economic price adjustment otherwise allowed under the Contract.

THE UNDERSIGNED CONTRACTING PARTIES do hereby certify that: (1) the services specified above are necessary and essential for activities that are properly within the statutory functions and programs of the affected agencies of State Government; (2) the proposed arrangements serve the interest of efficient and economical administration of the State Government, and (3) the services, supplies or materials contracted for are not required by Section 21 of Article 16 of the Constitution

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of Texas to be supplied under contract given to the lowest responsible bidder.

PERFORMING AGENCY further certifies that it has the authority to contract for the above services by authority granted in Article 4413 (32e) V.C.S. and Chapter 105 of the Texas Education Code.

RECEIVING AGENCY further certifies that it has the authority to perform the services contracted for by authority granted in Article 4413 (32e) V.C.S.

7.0 Points of Contact

Dr. Schonna R. Manning Florida International University - Biscayne Bay Campus 3000 NE 151st Street, MSB 250B/212 North Miami, Florida 33181, USA <u>schonna.manning@fiu.edu</u>

Brent Bellinger, Ph.D. City of Austin – Watershed Protection 505 Barton Springs Rd., Suite 1100 Austin, TX 78704 512-974-2717 Brent.Bellinger@austintexas.gov

FLORIDA INTERNATIONAL UNIVERSITY PROPOSAL BUDGET SHEET													
Principal Investigator/Project Director: Schonna R. Manning Agency: City of Austin Budget Period Start Date: 01/01/23 Budget Period End Date: 12/31/23 Period/Year #: 1													
Level 3	Budget Period Start Dates	01/01/23		Budget Period	End Date:					Pe	riod/ rear #:	┼──	I
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		771500-771549	1.	N/A Other Professionals						, c		\$	-
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	P77156 - Fringe									FRINGE BEN	IEFITS (S77100):		50,959.31
	P71121 - Domestic Travel	Object Code 711211-711226			PENSES - IN A	LPHABETIC		R BY BUDGI	ET CATEGORY			T	
	P71123 - Foreign Travel	711251-711286 711230-711239	1 2	Travel - Domestic Travel - Foreign								\$ \$	-
	P71119 - Long Distance Telephone Calls P77300 - Materials and Supplies	711190-711199 773600-773699	3 4	Long Distance Telephone Calls Educational & Lab Supplies								\$ \$	-
		773500-773599 773900-773910	5	Motor Fuel & Lubricants								\$	-
		773912 -773999 773911	6 7	6 Other Materials & Supplies 5 7 Recharge Center Charges 5								\$ \$	16,000.00 -
		773100-773199 711021-711031	8 9	Printing & Reproduction Freight								\$ \$	776.28
	P71190 - Other Operating Expenses	711360-711399 773001-773002	10	Insurance Miscellaneous Expenses (Other)								\$	-
		711075-711077 711900-711989	11	*** Must Provide Amount & Associated Details Below ***							\$	-	
S71100 - Operating Expenses		711902	12								\$	-	
	P71100 - Participant Payments P71150 - Patient Care Cost	711991 711510	13 14	Participant Payments Patient Care Costs								\$ \$	-
		772101-772102 772104-772126											
	P77200 - Professional Fees	772131-772132 772141 772151-772157											
	P77210 - Consultants	772301-772303 772103	15 16	Professional Fees Consulting Services/Agreements								\$	-
	P71140 - Rent Expense Other Than Buildings P76100 - Repairs and Maintenance	711401-711406 761001-761006	17 18	Rent Expense Other Than Buildings Repairs & Maintenance								\$	-
	P76830 - Scholarships P76830 - Stipends	768201-768211 768301	19 20	Scholarships Stipends								\$ \$	-
	p								TOTAL OPE	RATING EXPE	ENSES (S71100):	\$	16,776.28
	P77295 - Subcontractor Amounts over \$25K	772952		*** Subcontractor Budge	ets Must Be Ent	ered in The	e 'Project	Subcontracto	or Budgets' Wo	orksheet ***			
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	P7729U - Subcontractor Amounts under \$25K	nounts under \$25K 772951 *** Subcontractor Budgets Must Be Entered in The 'Project Subcontractor Budgets' Worksheet *** Subcontractor Budgets Must Be Entered in The 'Project Subcontractor Budgets' Worksheet ***									_		
	P76840 - Tuition	768400-768409	23	Tuition Payments								\$	-
Restricted Expenses - In alphabetical order by budget category. RED Items are part of F&A and are normally not allowed on Federal Grants. RED items will require a DIRECT CHARGE EXEMPTION FORM.													
S7710R - Restricted Operating Expenses (Restricted Items) \$0.00	P77220 - Advertising Services P72110 - Books & Film	772201-772204	24	Advertising Services								\$	-
	P77320 - Building and Construction	721101-721103 773201-773212	25 26	Books & Films Building & Construction Colluder Bhance								⇒ \$	-
	P71117 - Cellular Phones P77218 - Construction Services P77292 - Data Proceeding Supplies	711171 772181-772185	27 28	Cellular Phones Construction Services Data December Supplies								⇒ \$	-
	P77382 - Data Processing Supplies P77381 - Hazardous Chemicals/Waste	773821 773810	29 30	Data Processing Supplies Hazardous Chemicals/Waste								\$	-
	P77380 - Food Products P71116 - Local Telephone Calls	773801 711196	31 32	Food Products Local Telephone Calls								\$ \$	-
	P71145 - Memberships & Subscriptions P71130 - Moving Expenses	711451-711454 711301-711304	33 34	Membership & Subscriptions Moving Expenses								\$ \$	-
	P77330 - Office Supplies P71101 - Postage	773301-773305 711011-711013	35 36	Office Supplies/Computer Supplies Postage								\$	-
	P71144 - Rent Expense Buildings P71118-Telephone Equipment	711440 711180-711189	37 38	Rent Expense Buildings Telephone Equipment								\$ \$	-
TOTAL RESTRICTED OPER D. PERMANENT EQUIPMENT & BOOKS OR OTHER LIBRARY RESOURCES (list item and dollar)									RATING EXPE	NSES (S7710R):	\$	-	
S72120 - Equipment &			U. P	amount for each item e			(iiist item	and dollar)	•				
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S75700 - Facilities &				E. INDIREC	T COSTS				TOTAL	DIRECT COST	S (A through D):	\$	67,735.59
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				G. TOTAL DIRECT AND INDIRE	CT COSTS						,,	. ↓ \$	99,910.00